

Biosynthesis of Silver nanoparticles and Antibacterial activity using fungi *Aspergillus niger*

Madhumita Ghosh Dastidar, Sitrarasi.R and Razia.M*

Department of Biotechnology, Mother Teresa Women's University, Kodaikanal-624102

Abstract: An important area of research in nanotechnology is the biosynthesis of silver nanoparticles which are found to have wide applications. Micro-organisms and plant extracts have received attention for the biosynthesis of nanoparticles. A novel approach for the green synthesis of silver nanoparticles (AgNPs) from aqueous solution of AgNO₃ using fungus culture is reported in this work. Several fungi were isolated from soil among which *Aspergillus niger* was found to be most promising organism which were able to convert AgNO₃ solution to silver nanoparticles extracellularly. The synthesis was observed within 24h, and AgNPs showed characteristic absorbance around 410 nm. Spherical nanoparticles of size 50-100 nm were observed in transmission electron microscopy. The AgNPs showed highly potent antimicrobial activity against Gram-positive, Gram-negative bacteria. It may be concluded from this experiment that silver nanoparticles obtained from fungus are very significant and indicate that the synthesized silver nanoparticles may have an important advantage over conventional antibiotics.

Key Words: *Aspergillus niger*, Nanoparticles, Antibacterial activity, SEM.

Introduction

Nanotechnology has emerged rapidly during the past few years in a broad range of product domains. It provides opportunities for the development of materials, including those for medical applications, where many other conventional techniques may reach their limits. Nanotechnology represents the design, production and application of materials at atomic, molecular and macromolecular scales (Koper et al., 2002). The problem with most of the chemical and the physical methods of Nano molecule production is that they are extremely expensive and also involve the use of toxic, hazardous chemicals, which may pose potential environmental and biological risks. Pharmaceutical nanoparticles are solid, submicron-sized (1 to 100 nm in diameter) drug carriers that may or may not be biodegradable. Metal nanoparticles have attained a great importance due to their unique features such as catalytic, magnetic, optical and electrical properties (Alagumuthu and Kirubha, 2012). They have distinct or completely new properties when compared to their bulk counterpart, which draws the entire scientific research to focus on their synthesis (Bhattacharya and Rajinder, 2005).

Several metal nanoparticles such as silver, copper, gold, iron, etc. have been explored so far. Silver nanoparticles has major applications including use as catalysts, as optical sensor, in textile engineering, in electronics and most importantly in the medical field as a bactericidal and as a therapeutic agent (Kuhn et al., 2003). There is a need for an environmentally and economically feasible way to synthesize these nanoparticles. Biosynthesis of silver nanoparticles mostly involves reduction/oxidation reactions (Mandal et al., 2001). It is the microbial enzymes or the plant phytochemicals with antioxidant or reducing properties that act on the respective compounds and give the desired nanoparticles. In comparison with bacteria, fungi can produce larger amounts of nanoparticles as they can secrete larger amounts of proteins which directly translate to higher productivity of nanoparticles (Jiang et al., 2006). The mechanism of silver nanoparticle production by fungi occurs by trapping of Ag⁺ ions at the surface of the fungal cells and the subsequent reduction of the silver ions by the enzymes present in the fungal system (Kalishwaralal et al., 2008). The extracellular enzymes like nitrate reductase along with naphthoquinones and anthraquinones are said to facilitate the reduction.

Materials and Methods

Isolation of the Fungi

The garden soil sample was collected and was serially diluted and through pour plate technique *Aspergillus niger* was isolated in Sabouraud's agar by incubating the Petri plates at room temperature. The surface of the medium was filled with fungal mycelium after a week. The mycelium was observed under microscope using lacto phenol cotton blue. After microscopic observation and identification through morphological characteristics the fungus was sub-cultured and maintained in Sabouraud's agar.

Biosynthesis of silver nanoparticles using *A. niger*

Various concentrations of silver nitrate solutions (0.5M, 0.25M, and 0.125M) were prepared using double distilled water and were membrane filtered. 100ml Sabouraud's broth media was sterilized and inoculated with *A. niger* as well as silver nitrate solutions in various flasks. The conical flasks were incubated at room

temperature (Mohanpuria et al., 2008). The flask showing color changes from 24 hours onwards, by 48 hours it changed from pale yellow to brown and their color remained stable without any stabilizing agent.

The mat of fungal mycelium was separated from the broth by filtration. The silver nanoparticle solution thus obtained was centrifuged at 12,000 rpm for 15 min, after which the pellet was redispersed in deionized water and passed through membrane filter to get rid of impurities.

Characterization of Silver Nanoparticles

The silver nanoparticles were characterized by UV-Vis spectroscopy, one of the most widely used techniques for structural characterization of silver nanoparticles (Molpeceres et al., 2000). Scanning electron microscopy (SEM) was used for morphological examination by direct visualization. To gain further insight into the features of the silver nanoparticles, analysis of the sample was performed using EDAX techniques (Klaus et al., 1999).

Anti-bacterial activity analysis of Silver AgNPs

The antimicrobial activity of the silver nanoparticles were analysed through Well Diffusion method. Sterilized plates of Muller-Hinton Agar were prepared with a well of about 1 cm diameter to which multiple Antibiotic resistant *Pseudomonas aeruginosa* and *Bacillus subtilis* were swabbed. To the well 50 µl of AgNPs were added and incubated at 37°C for about 48 hours.

Results and Discussion

The absorption spectrum (Fig.1) of the yellowish-brown silver nanoparticle solution prepared showed a surface plasmon absorption band with a maximum of 440 nm, indicating the presence of Ag nanoparticles.

Scanning Electron Microscope (SEM)

The silver nanoparticles were observed at different magnifications. The shape obtained of AgNPs was spherical and the sizes of nanoparticles were 20-30 nm. The nanoparticles were uniformly distributed without significant agglomeration.

EDAX Analysis of AgNPs

The peak at 3.0 keV corresponds to the binding energies of AgNPs. The percentage of Ag and Oxygen significantly showed the production of AgNPs (Fig.2).

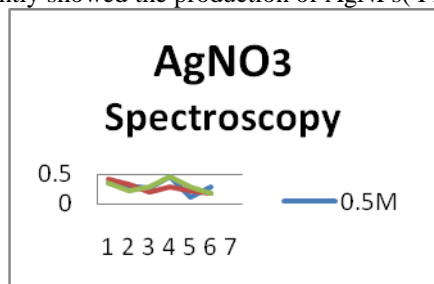


Fig.1 UV-Visible Absorption Spectrum

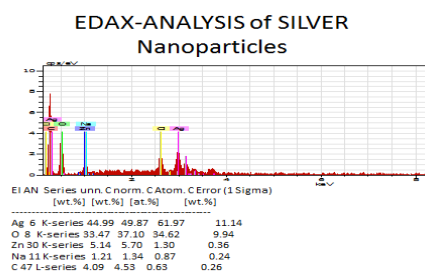


Fig.2 EDAX Analysis of AgNPs

Antimicrobial activity of AgNPs

The diameter of zone of inhibition of AgNPs was more against *P. aeruginosa* compared to *B. subtilis* (Table 1).

Concentration of AgNPs	Diameter of Zone of Inhibition (mm)	
	<i>Pseudomonas aeruginosa</i>	<i>Bacillus subtilis</i>
0.5M	35	18
0.25M	26	20
0.125m	24	20
AgNO ₃ Solution	16	15

Table 1: Antibacterial activity of AgNPs against *Pseudomonas aeruginosa* and *Bacillus subtilis*

Conclusion

A stable and eco-friendly metallic nanoparticle is needed to be developed in the field of nanotechnology. Since fungi contain enzymes and proteins as reducing agents, these can convert metal salts to metal nanoparticles faster. Fungal biomass normally grows faster than those of bacteria (Sastry et al., 2003)

under the same conditions. Although synthesis of metal nanoparticles by bacteria is prevalent, their synthesis by fungi is more advantageous because their mycelia offer a large surface area for interaction. The obtained silver nanoparticles were analyzed using UV-Vis, SEM and EDAX techniques. The results confirmed the reduction of silver nitrate to silver nanoparticles with high stability with an average size between 20–30nm. In this study the AgNPs were effective against multiple antibiotic resistant *Pseudomonas aeruginosa* (Madhumita and Razia, 2016). The nanoparticles and antibiotics had a synergistic effect against the same bacteria. These nanoparticles could be of immense use to control multiple drug resistant pathogens in medical fields. These nanoparticles can be used in textile industries because of efficient dye reduction capacity.

References

- [1]. Ahmad A, Mukherjee P, Senapati S, Mandal D, Khan MI, Kumar R, Sastry M (2003) Extracellular biosynthesis of silver nanoparticles using the fungus *Fusarium oxysporum*. *Colloids Surf. B Biointerfaces*, 28: 313–318.
- [2]. Bhainsa KC, D'Souza SF (2006) Extracellular biosynthesis of silver nanoparticles using the fungus *Aspergillus fumigatus*. *Coll Surf B Biointerfaces* 47:160–164
- [3]. Bhattacharya D, Rajinder G (2005) Nanotechnology and potential of microorganisms. *Crit Rev Biotechnology* 25: 199–204.
- [4]. Binupriya AR, Sathishkumar M, Yun SI (2010) Myco-crystallization of silver ions to nanosized particles by live and dead cell filtrates of *Aspergillus oryzae* var. *viridis* and its bactericidal activity toward *Staphylococcus aureus* KCCM 12256. *IndEngChem Res* 49: 852–858.
- [5]. Chen JC, Lin ZH, Ma XX (2003) Evidence of the production of silver nanoparticles via pretreatment of *Phoma* sp.3.2883 with silver nitrate. *LettApplMicrobiol* 37:105–108.
- [6]. Alagumuthu G, Kirubha R (2012) Green Synthesis of silver nanoparticles using *Cissus quadrangularis* plant extract their antibacterial activity University, *International Journal of Nanomaterials and Biostructures*, 2:30–33.
- [7]. Husen A, Siddiqi KS (2014) Plants and microbes assisted selenium nanoparticles: characterization and application. *J Nanobiotechnol* 12:28.
- [8]. Irvani S (2014) Bacteria in nanoparticle synthesis: current status and future prospects. *IntSchol Res Notic* 18:3593.
- [9]. Jiang H, Moon K, Zhang Z, Pothukuchi S, Wong CP (2006) Variable frequency microwave synthesis of silver nanoparticles. *J.Nanopart. Res.* 8: 117–124.
- [10]. Jores K, Mehnert W, Drecusler M, Bunyes H, Johan , Mader K (2004) Investigation on the stricter of solid lipid nanoparticles and oil-loaded solid nanoparticles by photon correlation spectroscopy, fieldflowfractionation and transmission electron microscopy. *J Control*; 17: 217– 227.
- [11]. Kalishwaralal K, Deepak V, Ramkumarpandian S, Nellaiah H, Sangiliyandi G (2008) Extracellular biosynthesis of silver nanoparticles by the culture supernatant of *Bacillus licheniformis*. *Mater. Lett.*, 62:4411–4413.
- [12]. Klaus T, Joerger R, Olsson E, Granqvist CG (1999) “Silver-Based Crystalline Nanoparticles, Microbially Fabricated”, *J. Proc. Natl. Acad. Sci. USA*, 96, 13611–13614,
- [13]. Koper OB, Klabunde JS, Marchin GL, Klabunde KJ, Stoimenov P, Bohra L (2002) Nanoscale powders and formulations with biocidal activity toward spores and vegetative cells of *Bacillus* species, viruses, and toxins. *CurrMicrobiol* 44: 49–55.
- [14]. Kuhn KP, Chaberny IF, Maschholder K, Stickler M, Benz VW, Sonntag HG, Erdinger L (2003) Disinfection of surfaces by photocatalytic oxidation with titanium dioxide and UVA light. *Chemosphere* 53: 71–77.
- [15]. Lee HJ, Yeo SY, Jeong SH (2003) Antibacterial effect of nanosized silver colloidal solution on textile fabrics. *J Mater Sci* 38: 2199–2204.
- [16]. Madhumita G D , Razia M (2016) Isolation of Multiple Drug Resistant bacteria from hospital environment, *Int.J.Microbiol.App.Sci.*, 5:48–53.
- [17]. Mandal S, Arumugam S, Pasricha R, Sastry M (2001) Silver nanoparticles of variable morphology synthesized in aqueous foams as novel templates. *Bull. Mater. Sci.*, 28: 503–510.
- [18]. Mishra A, Tripathy S, Yun SI (2011) Bio-synthesis of gold and silver nanoparticles from *Candida guilliermondii* and their antimicrobial effect against pathogenic bacteria. *J Nanosci Nanotechnol* 1:243–248
- [19]. Mohanpuria P, Rana KN, Yadav SK (2008) Biosynthesis of nanoparticles: technological concepts and future applications. *J. Nanopart. Res.* 10: 507–517.
- [20]. Molpeceres J, Aberturas MR, Guzman M (2000) Biodegradable nanoparticles as a delivery system for cyclosporine: preparation and characterization. *J Microencapsul.* 17: 599–614.

- [21]. Sastry M, Ahmad A, Khan MI, Kumar R (2003) Biosynthesis of metal nanoparticles using fungi and actinomycete. *Cur Sci* 85:162–170
- [22]. Singh R, Shedbalkar UU, Wadhwani SA, Chopade BA (2015) Bacteriogenic silver nanoparticles: synthesis, mechanism, and applications. *App MicrobiolBiotechnol* 99:4579–4593
- [23]. Vahabi K, Mansoori GA, Karimi S (2011) Biosynthesis of silver nanoparticles by fungus *Trichoderma reesei* (a route for large-scale production of AgNPs). *Insci J* 1:65–79
- [24]. Vigneshwaran N, Ashtaputre NM, Varadarajan PV, Nachane, R, Paralikar, KM, Balasubramanya, RH (2007) “Biological Synthesis of Silver Nanoparticles Using the Fungus *Aspergillus flavus*”, *Mater. Lett.*, 61, 1413-1418,
- [25]. Vilchis-Nestor AR, Sanchez-Mendieta V, Camacho-Lopez M, Gomez-Espinosa RM, Camacho-Lopez MA, et al. (2008) Solvent less synthesis and optical properties of Au and Ag nanoparticles using *Camellia sinensis* extract. *Mater Lett* 62: 3103-3105.
- [26]. Zhang J, Chen P, Sun C, Hu X (2004) Sonochemical synthesis of colloidal silver catalysts for reduction of complexing silver in DTR system. *ApplCatal A Gen* 266: 49-54.